

Mouse LTF/LF (Lactoferrin) ELISA Kit

Catalog No: BGT-KET-14504

Size: 96 assays

Intended use

This ELISA kit applies to the in vitro quantitative determination of Mouse LTF/LF concentrations in serum, plasma and other biological fluids.

Character

Item	
Sensitivity	15.2 pg/mL
Detection Range	31.25-2000 pg/mL
Specificity	This kit recognizes Mouse LTF/LF in samples. No significant cross-reactivity or interference between Mouse LTF/LF and analogues was observed
Repeatability	Coefficient of variation is < 10%

Test principle

This ELISA kit uses the Sandwich-ELISA principle. The micro ELISA plate provided in this kit has been pre-coated with an antibody specific to Mouse LTF/LF. Samples (or Standards) are added to the micro ELISA plate wells and combined with the specific antibody. Then a biotinylated detection antibody specific for Mouse LTF/LF and Avidin-Horseradish Peroxidase (HRP) conjugate are added successively to each micro plate well and incubated. Free components are washed away. The substrate solution is added to each well. Only those wells that contain Mouse LTF/LF, biotinylated detection antibody and Avidin-HRP conjugate will appear blue in color. The enzyme-substrate reaction is terminated by the addition of stop solution and the color turns yellow. The optical density (OD) is measured spectrophotometrically at a wavelength of 450 ± 2 nm. The OD value is proportional to the concentration of Mouse LTF/LF. You can calculate the concentration of Mouse LTF/LF in the samples by comparing the OD of the samples to the standard curve.

Kit components & Storage

The kit can be stored at 2-8°C for 3 months. If the kit is used after 3 months, store the items separately according to the following conditions once the kit is received.

Item	Specifications	Storage
Micro ELISA Plate (Dismountable)	8 wells × 12 strips	-20°C, up to expiry date (6 months)
Reference Standard	2 vials	
Concentrated Biotinylated Detection Ab(50×)	1 vial, 120 µL	
Concentrated HRP Conjugate (50×)	1 vial, 120 µL	
Reference Standard & Sample Diluent	3 vials, 20 mL	2-8°C, up to expiry date (6 months)
Biotinylated Detection Ab Diluent	1 vial, 14 mL	
HRP Conjugate Diluent	1 vial, 14 mL	
Concentrated Wash Buffer(25×)	1 vial, 30 mL	
Substrate Reagent	1 vial, 10 mL	
Stop Solution	1 vial, 10 mL	
Plate Sealer	5 pieces	
Product Description	1 copy	
Certificate of Analysis	1 copy	

Note: Concentrated HRP Conjugate(100×) and Substrate Reagent should be stored away from light.

All reagent bottle caps must be tightened to prevent evaporation and microbial pollution. The volume of reagents in partial shipments is a little more than the volume marked on the label, please use accurate measuring equipment instead of directly pouring into the vial(s).

Other supplies required

Microplate reader with 450 nm wavelength filter
High-precision transfer pipette, EP tubes and disposable pipette tips
Incubator capable of maintaining 37°C
Deionized or distilled water
Absorbent paper
Loading slot

Sample collection

Serum: Allow samples to clot for 1 hour at room temperature or overnight at 2-8°C before centrifugation for 20 min at 1000 ×g at 2-8°C. Collect the supernatant to carry out the assay.

Plasma: Collect plasma using EDTA-Na₂ as an anticoagulant. Centrifuge samples for 15 min at 1000 ×g at 2-8°C within 30 min of collection. Collect the supernatant to carry out the assay.

Tissue homogenates: It is recommended to get detailed references from the literature before analyzing different tissue types. For general information, hemolyzed blood may affect the results, so the tissues should be minced into small pieces and rinsed in ice-cold PBS (0.01M, pH=7.4) to remove excess blood thoroughly. Tissue pieces should be weighed and then homogenized in PBS (tissue weight (g): PBS (mL) volume=1:9) with a glass homogenizer on ice. To further break down the cells, you can sonicate the suspension with an ultrasonic cell disrupter or subject it to freeze-thaw cycles. The homogenates are then centrifuged for 5-10 min at 5000 ×g at 2-8°C to get the supernatant.

Cell lysates: For adherent cells, gently wash the cells with moderate amount of pre-cooled PBS and dissociate the cells using trypsin. Collect the cell suspension into a centrifuge tube and centrifuge for 5 min at 1000 ×g. Discard the medium and wash the cells 3 times with pre-cooled PBS. For each 1 ×10⁶ cells, add 150-250 μL of pre-cooled PBS to keep the cells suspended. Repeat the freeze-thaw process several times or use an ultrasonic cell disrupter until the cells are fully lysed. Centrifuge for 10 min at 1500 ×g at 2-8°C. Remove the cell fragments, collect the supernatant to carry out the assay.

Cell culture supernatant or other biological fluids: Centrifuge samples for 20 min at 1000 ×g at 2-8°C. Collect the supernatant to carry out the assay.

Recommended reagents for sample preparation: PMSF Protease Inhibitor, 0.25% Trypsin Solution.

Note

■ Note for kit

- 1) For research use only. Not for use in diagnostic procedures.
- 2) Please wear lab coats, eye protection and latex gloves for protection. Please perform the experiment following the national security protocols of biological laboratories, especially when detecting blood samples or other bodily fluids.
- 3) A freshly opened ELISA plate may appear a water-like substance, which is normal and will not have any impact on the experimental results. Return the unused wells to the foil pouch and store according to the conditions suggested in the above table.
- 4) Do not reuse the reconstituted standard, biotinylated detection Ab working solution, HRP conjugate working solution. The unspent undiluted concentrated biotinylated detection Ab (50×) and other stock solutions should be stored according to the storage conditions in the above table.
- 5) The microplate reader should be able to be installed with a filter that can detect the wave length at 450 ± 2 nm. The optical density should be within 0-3.5. Follow the Instructions of the Microplate Reader for set-up and preheat it for 15 min before OD measurement.
- 6) Do not mix or substitute reagents with those from other lots or sources.
- 7) Change pipette tips in between adding of each standard level, between sample adding and between reagent adding. Also, use separate reservoirs for each reagent.
- 8) The kit should not be used beyond the expiration date on the kit label.

■ Note for sample

- 1) Tubes for blood collection should be disposable and be non-endotoxin. Samples with high hemolysis or much lipid are not suitable for ELISA assay.
- 2) Samples should be assayed within 7 days when stored at 2-8°C, otherwise samples must be divided up and stored at -20°C (≤ 1 month) or -80°C (≤ 3 months). Avoid repeated freeze-thaw cycles. Prior to assay, the frozen samples should be slowly thawed and centrifuged to remove precipitates.
- 3) Please predict the concentration before assaying. If the sample concentration is not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.
- 4) If the sample type is not included in the manual, a preliminary experiment is suggested to verify the validity.
- 5) If a lysis buffer is used to prepare tissue homogenates or cell lysates, there is a possibility of causing a deviation due to the introduced chemical substance.
- 6) Some recombinant protein may not be detected due to a mismatching with the coated antibody or detection antibody.

Dilution method

It is suggested to dilute normal serum/plasma samples at 500~5000 fold.

Due to individual differences, please estimate the concentration range of the sample in advance, and conduct a preliminary test to determine the appropriate dilution ratio of the sample.

If your test sample needs dilution, please refer to the dilution method as follows:

For 100 fold dilution: One-step dilution. Add 5 μ L sample to 495 μ L sample diluent to yield 100 fold dilution.

For 1000 fold dilution: Two-step dilution. Add 5 μ L sample to 95 μ L sample diluent to yield 20 fold dilution, then add 5 μ L 20 fold diluted sample to 245 μ L sample diluent, after this, the neat sample has been diluted at 1000 fold successfully.

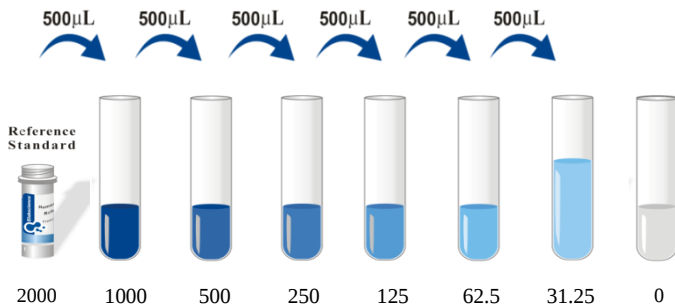
For 100000 fold dilution: Three-step dilution. Add 5 μ L sample to 195 μ L sample diluent to yield 40 fold dilution, then add 5 μ L 40 fold diluted sample to 245 μ L sample diluent to yield 50 fold dilution, and finally add 5 μ L 2000 fold diluted sample to 245 μ L sample diluent, after this, the neat sample has been diluted at 100000 fold successfully.

Reagent preparation

1. Bring all reagents to room temperature (18-25 $^{\circ}$ C) before use. If the kit will not be used up in one assay, please only take out the necessary strips and reagents for present experiment, and store the remaining strips and reagents at required condition.
2. **Wash Buffer:** Dilute 30 mL of Concentrated Wash Buffer with 720 mL of deionized or distilled water to prepare 750 mL of Wash Buffer. Note: if crystals have formed in the concentrate, warm it in a 40 $^{\circ}$ C water bath and mix it gently until the crystals have completely dissolved. For same day use only.
3. **Standard working solution:** Centrifuge the standard at 10,000 $\times$ g for 1 min. Add 1 mL of Reference Standard & Sample Diluent, let it stand for 10 min and invert it gently several times. After it dissolves fully, mix it thoroughly with a pipette. This reconstitution produces a working solution of 2000 pg/mL (or add 1 mL of Reference Standard & Sample Diluent, let it stand for 1-2 min and then mix it thoroughly with a vortex meter of low speed. Bubbles generated during vortex could be removed by centrifuging at a relatively low speed). Then make serial dilutions as needed. The recommended dilution gradient is as follows: 2000, 1000, 500, 250, 125, 62.5, 31.25, 0 pg/mL.

Dilution method: Take 7 EP tubes, add 500 μ L of Reference Standard & Sample Diluent to each tube. Pipette 500 μ L of the 2000 pg/mL working solution to the first tube and mix up to produce a 1000 pg/mL working solution. Pipette 500 μ L of the solution from the former tube into the latter one according to this step. The illustration on the next page is for reference. Note: the last tube is regarded as a blank. Don't pipette

solution into it from the former tube. Gradient diluted standard working solution should be prepared just before use.



4. **Biotinylated Detection Ab working solution:** Calculate the required amount before the experiment ($50\mu\text{L}/\text{well}$). In preparation, slightly more than calculated should be prepared. Centrifuge the Concentrated Biotinylated Detection Ab at $800\times g$ for 1 min, then dilute the $50\times$ Concentrated Biotinylated Detection Ab to $1\times$ working solution with Biotinylated Detection Ab Diluent (Concentrated Biotinylated Detection Ab: Biotinylated Detection Ab Diluent= 1: 49). The working solution should be prepared just before use.
5. **HRP Conjugate working solution:** HRP Conjugate is HRP conjugated avidin. Calculate the required amount before the experiment ($50\mu\text{L}/\text{well}$). In preparation, slightly more than calculated should be prepared. Centrifuge the Concentrated HRP Conjugate at $800\times g$ for 1 min, then dilute the $50\times$ Concentrated HRP Conjugate to $1\times$ working solution with HRP Conjugate Diluent (Concentrated HRP Conjugate: HRP Conjugate Diluent= 1: 49). The working solution should be prepared just before use.

Assay procedure

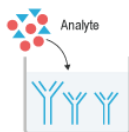
1. Determine wells for **diluted standard, blank** and **sample**. Add 25 μL each dilution of standard, blank and sample into the appropriate wells (It is recommended that all samples and standards be assayed in duplicate. It is recommended to determine the dilution ratio of samples through preliminary experiments or technical support recommendations). Cover the plate with the sealer provided in the kit. Incubate for 90 min at 37°C. Note: solutions should be added to the bottom of the micro ELISA plate well, avoid touching the inside wall and causing foaming as much as possible.
2. Decant the liquid from each well, do not wash. Immediately add 50 μL of **Biotinylated Detection Ab working solution** to each well. Cover the plate with a new sealer. Incubate for 1 hour at 37 °C.
3. Decant the solution from each well, add 350 μL of **wash buffer** to each well. Soak for 1 min and aspirate or decant the solution from each well and pat it dry against clean absorbent paper. Repeat this wash step 3 times. Note: a microplate washer can be used in this step and other wash steps. Make the tested strips in use immediately after the wash step. Do not allow wells to be dry.
4. Add 50 μL of **HRP Conjugate working solution** to each well. Cover the plate with a new sealer. Incubate for 30 min at 37 °C.
5. Decant the solution from each well, repeat the wash process for 5 times as conducted in step 3.
6. Add 50 μL of **Substrate Reagent** to each well. Cover the plate with a new sealer. Incubate for about 15 min at 37 °C. Protect the plate from light. Note: the reaction time can be shortened or extended according to the actual color change, but not more than 30 min. Preheat the Microplate Reader for about 15 min before OD measurement.
7. Add 25 μL of **Stop Solution** to each well. Note: adding the stop solution should be done in the same order as the substrate solution.
8. Determine the optical density (OD value) of each well at once with a micro-plate reader set to 450 nm.

Calculation of results

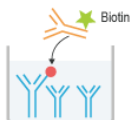
Average the duplicate readings for each standard and samples, then subtract the average zero standard optical density. Plot a four parameter logistic curve on log-log axis, with standard concentration on the x-axis and OD values on the y-axis.

If the OD of the sample surpasses the upper limit of the standard curve, you should re-test it with an appropriate dilution. The actual concentration is the calculated concentration multiplied by the dilution factor.

Assay Procedure Summary



1. Add 25 μ L standard or sample to the wells.
Incubate for 90 min at 37°C



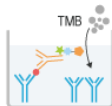
2. Discard the liquid, immediately add 50 μ L Biotinylated Detection Ab working solution to each well. Incubate for 60 min at 37°C



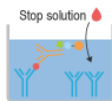
3. Aspirate and wash the plate for 3 times



4. Add 50 μ L HRP conjugate working solution.
Incubate for 30 min at 37°C. Aspirate and wash the plate for 5 times



5. Add 50 μ L Substrate Reagent. Incubate for 15 min at 37°C



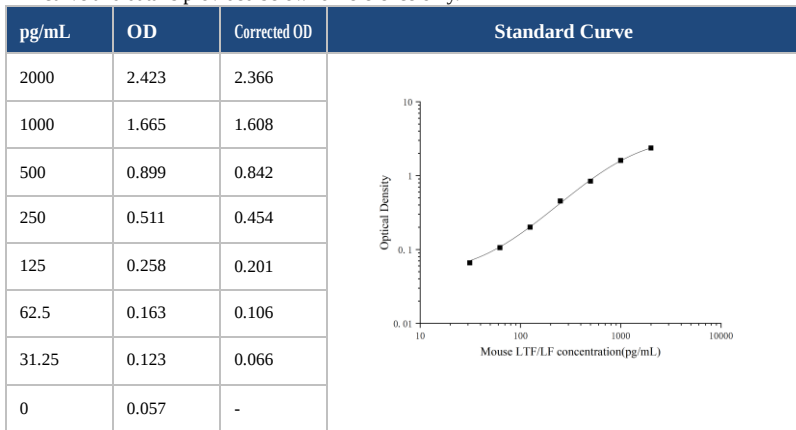
6. Add 25 μ L Stop Solution



7. Read the plate at 450nm immediately.
Calculation of the results

Typical data

As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test. Typical standard curve and data is provided below for reference only.



Performance

■ Precision

Intra-assay Precision (Precision within an assay): 3 samples with low, mid range and high level Mouse LTF/LF were tested 20 times on one plate, respectively.

Inter-assay Precision (Precision between assays): 3 samples with low, mid range and high level Mouse LTF/LF were tested on 3 different plates, 20 replicates in each plate, respectively.

	Intra-assay Precision			Inter-assay Precision		
Sample	1	2	3	1	2	3
n	20	20	20	20	20	20
Mean(pg/mL)	109.54	185.35	984.48	111.94	169.18	1081.09
Standard deviation	5.78	11.12	35.05	6.38	9.25	39.89
CV (%)	5.28	6	3.56	5.7	5.47	3.69

■ Recovery

The recovery of Mouse LTF/LF spiked in three different levels in samples throughout the range of the assay was evaluated in various matrices.

Sample Type	Range (%)	Average Recovery (%)
Serum (n=8)	96-109	101
EDTA plasma (n=8)	92-108	100
Cell culture media(n=8)	97-109	102

■ Linearity

Samples were spiked with high concentrations of Mouse LTF/LF and diluted with Reference Standard & Sample Diluent to produce samples with values within the range of the assay.

		Serum (n=5)	EDTA plasma (n=5)	Cell culture media(n=5)
1:2	Range (%)	91-105	87-103	85-97
	Average (%)	98	94	92
1:4	Range (%)	90-102	86-100	85-97
	Average (%)	97	92	92
1:8	Range (%)	91-104	88-102	88-101
	Average (%)	98	95	94
1:16	Range (%)	92-106	88-101	82-95
	Average (%)	100	95	88

Declaration

1. Limited by current conditions and scientific technology, we can't conduct comprehensive identification and analysis on all the raw material provided. So there might be some qualitative and technical risks for users using the kit.
2. This assay is designed to eliminate interference by factors present in biological samples. Until all factors have been tested in the ELISA immunoassay, the possibility of interference cannot be excluded.
3. The final experimental results will be closely related to the validity of products, operational skills of the operators, the experimental environments and so on. We are only responsible for the kit itself, but not for the samples consumed during the assay. The users should calculate the possible amount of the samples used in the whole test. Please reserve sufficient samples in advance.
4. To get the best results, please only use the reagents supplied by the manufacturer and strictly comply with the instructions.
5. Incorrect results may occur because of incorrect operations during the reagents preparation and loading, as well as incorrect parameter settings of the Micro-plate reader. Please read the instructions carefully and adjust the instrument prior to the experiment.
6. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
7. Every kit has strictly passed QC test. However, results from end users might be inconsistent with our data due to some variables such as transportation conditions, different lab equipment, and so on. Intra-assay variance among kits from different batches might arise from the above reasons too.
8. Kits from different manufacturers or other methods for testing the same analyte could bring out inconsistent results, since we haven't compared our products with those from other manufacturers.
9. The kit is designed for research use only, we will not be responsible for any issues if the kit is applied in clinical diagnosis or any other related procedures.