

Eosin Staining Solution (Alcohol-Based, 0.5%)

Cat #: A-CSH269

Size: 100mL, 500mL

Storage: 10-30 °C, Avoid exposure to light (1 year)

Product Description

Eosin, also known as erythrosin, is a xanthene dye belonging to the class of synthetic dyes. It is a pink or peach-colored powder and is most suitable for staining in combination with hematoxylin to visualize the morphological structure of normal or pathological tissues. The eosin staining solution (alcohol-based, 0.5%) is prepared using high-quality alcohol-soluble eosin, offering a simple operation without the use of toxic reagents such as mercury or methanol. It is commonly used for staining tissue sections or cultured cells, and the cytoplasm appears pink or red after staining.

Usage

I. Sample Preparation

A. Paraffin Section Preparation:

1. Bake the slides for 30-60 minutes and dewax in xylene (I) for 5-10 minutes.
2. Dewax in xylene (II) for 5-10 minutes.
3. Wash in absolute ethanol for 1-5 minutes.
4. Wash in 95% ethanol for 1-5 minutes.
5. Wash in 75% ethanol for 1-5 minutes.
6. Rinse with tap water or distilled water.

B. Frozen Section Preparation:

1. No dewaxing is required. After fixation, rinse directly with distilled water for 2-3 minutes.

C. Cell Sample Preparation:

1. Fix with 4% paraformaldehyde for 10-20 minutes.
2. Rinse twice with tap water for 2 minutes each time.

3. Rinse twice with distilled water for 2 minutes each time.

II. Staining

1. Staining with eosin staining solution (adjusting the time based on staining results and requirements).
2. Rinse with 75%~85% ethanol for 30 seconds.

Note: If eosin counterstaining is performed after immunohistochemistry or other staining, refer to the above steps and perform eosin staining after the completion of the other staining procedure.

III. Dehydration, Clearing, and Mounting (optional)

1. Wash with 95% ethanol (I) for 0.5-2 minutes.
2. Dehydrate with 95% ethanol (II) for 2-5 minutes.
3. Dehydrate with absolute ethanol (I) for 2-5 minutes.
4. Dehydrate with absolute ethanol (II) for 2-5 minutes.
5. Clear with xylene (I) for 1 minute.
6. Xylene (II) clearing for 1 minute, followed by neutral resin mounting.
7. Observe under a microscope, and the cytoplasm should appear pink or red.

IV. Perform Other Staining:

1. For immunofluorescence staining or staining with fluorescent dyes such as Hoechst, wash twice with 70% ethanol after eosin staining.
2. Soak in PBS, saline, TBS, TBST, or other solutions used for immunostaining or fluorescent dye staining for 5 minutes.
- C. Perform immunofluorescence staining or staining with other fluorescent dyes.

Staining Results

Cytoplasm and fibers	Red
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Notes

1. If dehydration, clearing, and mounting are required, prepare xylene, neutral resin, or other mounting agents.

2. For your safety and health, please wear lab coat and disposable gloves while performing the procedure.

Disclaimer

The reagent is only used in the field of scientific research, not suitable for clinical diagnosis or other purposes.