Eosin Staining Solution (Aqueous, 5%)

Cat #: A-CSH265

Size: 100mL

Storage: 10-30 °C, Avoid exposure to light (1 year)

Product Description

Eosin, also known as erythrosin, is a xanthene dye belonging to the class of synthetic dyes. It is a pink or peach-colored powder and is most suitable for staining in combination with hematoxylin to visualize the morphological structure of normal or pathological tissues. The eosin staining solution (imported and water-soluble, 5%) utilizes imported eosin dye, offering a simple operation without the use of toxic reagents such as mercury or methanol. It can be used for staining tissue sections or cultured cells. It can be combined with immunofluorescence staining or immunohistochemistry staining. On one hand, it can be used for immunofluorescence staining or staining with other dyes after eosin staining, and on the other hand, it can be used for eosin counterstaining after immunohistochemistry staining. This staining solution can be reused until the staining results are considered unsatisfactory, and the cytoplasm appears pink or red after staining.

Usage

I. Sample Preparation

A. Paraffin Section Preparation:

1. Bake the slides for 30-60 minutes and dewax in xylene (I) for 5-10 minutes.

2. Dewax in xylene (II) for 5-10 minutes.

3. Wash in absolute ethanol for 1-5 minutes.

4. Wash in 95% ethanol for 1-5 minutes.

5. Wash in 75% ethanol for 1-5 minutes.

6. Rinse with tap water or distilled water.

B. Frozen Section Preparation:

1. No dewaxing is required. After fixation, rinse directly with distilled water for 2-3 minutes.

C. Cell Sample Preparation:

1. Fix with 4% paraformaldehyde for 10-20 minutes.

2. Rinse twice with tap water for 2 minutes each time.

3. Rinse twice with distilled water for 2 minutes each time.

II. Staining

1. Stain with eosin staining solution for 2-5 minutes (adjust the time based on staining results and requirements).

If direct observation is needed, wash twice with 70% ethanol. If dehydration, clearing, and mounting are required, follow the subsequent steps. After washing with 70% ethanol, proceed with dehydration, clearing, and mounting according to the subsequent steps.

Note: If used for counterstaining after immunohistochemistry or other staining methods, refer to the above steps and perform eosin staining after completing the other staining procedures.

III. Dehydration, Clearing, and Mounting (optional)

1. Wash with 95% ethanol (I) for 0.5-2 minutes.

2. Dehydrate with 95% ethanol (II) for 2-5 minutes.

3. Dehydrate with absolute ethanol (I) for 2-5 minutes.

4. Dehydrate with absolute ethanol (II) for 2-5 minutes.

5. Clear with xylene (I) for 1 minute.

6. Clear with xylene (II) for 1 minute and mount with neutral resin.

7. Observe under a microscope, and the cytoplasm should appear pink or red.

IV. Perform Other Staining:

1. For immunofluorescence staining or staining with fluorescent dyes such as Hoechst, wash twice with 70% ethanol after eosin staining.

2. Soak in PBS, saline, TBS, TBST, or other solutions used for immunostaining or fluorescent dye staining for 5 minutes.

C. Perform immunofluorescence staining or staining with other fluorescent dyes.

Staining Results

Cytoplasm and fibers	Red

Notes

1. If dehydration, clearing, and mounting are required, you will need to prepare xylene, neutral mounting medium, or other mounting agents. If the samples are paraffin sections, you will need to prepare 90% ethanol, absolute ethanol, and xylene.
2. For your safety and health, please wear laboratory attire and disposable gloves while performing the procedure.

Disclaimer

The reagent is only used in the field of scientific research, not suitable for clinical diagnosis or other purposes.