

Eosin Staining Solution (Alcohol-Based, 0.25%)

Cat #: A-CSH263

Size: 100mL, 500mL

Storage: 10-30 °C, Avoid exposure to light (1 year)

Product Description

Eosin, also known as erythrosin, is a xanthene dye belonging to the class of synthetic dyes. It is a pink or peach-colored powder and is commonly used in combination with hematoxylin for staining to visualize the morphological structure of normal or pathological tissues. Eosin staining solution (alcohol-soluble, 0.25%) is prepared using high-quality alcohol-soluble eosin, which is simple to use and does not require the use of toxic reagents such as mercury or methanol. It is commonly used for staining tissue sections or cultured cells, resulting in a pink or red cytoplasm after staining.

Usage

I. Sample Preparation

A. Paraffin Section Preparation:

1. Bake the slides for 30-60 minutes and dewax in xylene (I) for 5-10 minutes.
2. Dewax in xylene (II) for 5-10 minutes.
3. Wash in absolute ethanol for 1-5 minutes.
4. Wash in 95% ethanol for 1-5 minutes.
5. Wash in 75% ethanol for 1-5 minutes.
6. Rinse with tap water or distilled water.

B. Frozen Section Preparation:

1. No dewaxing is required. After fixation, rinse directly with distilled water for 2-3 minutes.

C. Cell Sample Preparation:

1. Fix with 4% paraformaldehyde for 10-20 minutes.

2. Rinse twice with tap water for 2 minutes each time.
3. Rinse twice with distilled water for 2 minutes each time.

II. Staining

1. Stain with eosin staining solution (adjust the time based on staining results and requirements).
2. Wash with 75-85% ethanol for 30 seconds.

Note: If used for counterstaining in immunohistochemistry or other staining methods, perform eosin staining after completing the other staining steps mentioned above.

III. Dehydration, Clearing, and Mounting (optional)

1. Wash with 95% ethanol (I) for 0.5-2 minutes.
2. Dehydrate with 95% ethanol (II) for 2-5 minutes.
3. Dehydrate with absolute ethanol (I) for 2-5 minutes.
4. Dehydrate with absolute ethanol (II) for 2-5 minutes.
5. Clear with xylene (I) for 1 minute.
6. Clear with xylene (II) for 1 minute and mount with neutral resin.
7. Observe under a microscope, and the cytoplasm should appear pink or red.

IV. Perform Other Staining:

1. For immunofluorescence staining or staining with fluorescent dyes such as Hoechst, wash twice with 70% ethanol after eosin staining.
2. Soak in PBS, saline, TBS, TBST, or other solutions used for immunostaining or fluorescent dye staining for 5 minutes.
- C. Perform immunofluorescence staining or staining with other fluorescent dyes.

Staining Results

Cytoplasm and fibers	Red
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Notes

1. If dehydration, clearing, and mounting are required, prepare xylene, neutral resin, or other mounting agents.
2. For your safety and health, wear lab coat and disposable gloves while performing the procedure.

Disclaimer

The reagent is only used in the field of scientific research, not suitable for clinical diagnosis or other purposes.